Bioremediation of V⁵⁺ and Ni²⁺ by indigenous *Marinobacter Goseongensis* isolated from South African Vanadium Mine Waste

Ilunga Каміка; Maggy NB Момва

Department of Environmental, Water and Earth Sciences, Faculty of Science, Tshwane University of Technology, Arcadia Campus, Private Bag X680, Pretoria 0001, South Africa. mombamnb@tut.ac.za

Abstract Study assessed the bioremediation of V⁵⁺ and Ni²⁺ in wastewater by an indigenous bacterial species. Bacterial species was isolated from mine water and characterized by sequencing. The isolate appeared to tolerate V⁵⁺ and Ni²⁺ separately at concentrations ranging between 650 to 700 mg/L and 250 and 300 mg/L (24h-LC₅), respectively. Removal of V⁵⁺ and Ni²⁺ in the media was inversely proportional to the test metal concentrations. *Marinobacter goseongensis* could remove more than 99.99 % of both metals at 50 mg/L at below the maximum permissible limit of 0.1 mg/L. This indigenous bacterial isolate can be used in V⁵⁺ and Ni²⁺ bioremediation.

Keywords Mine water, microbial diversity, *Marinobacter*, vanadium, nickel, pollution, bioremediation

Introduction

Mine water remains one of the major problems of concern, not only in South Africa, but also worldwide. This is due to its environmental, socio-economic and public health impacts (Oelofse 2009). In South Africa in particular, mining activities have a long history and have played a major role in both economic development and environmental pollution countrywide (Adler et al. 2007). Although significant progress has been made to address mine water management, environmental pollution due to the disposal of untreated mine water still remains a challenge. Microorganisms, due to their ubiquitousness, have been viewed as one of the best ways to deal with this problem. Due to their ability to survive, grow and reproduce in such harsh environments, an interest in microorganisms was aroused among researchers worldwide (Zhou et al. 2010). Nevertheless, their presence in extreme environments such as mine water affects their species diversity (Wang et al. 2011). Wang et al. (2011) have pointed out that extreme conditions can be defined by levels of environmental factors, the effects of which pose difficulties for the survival of specific

taxa or all taxa. In addition, both Johnson and Hallberg (2003) and Imarla et al. (2006) have also reported that a microbial community composition is largely bound to geochemical parameters such as pH and metal ion concentrations. As a result, microorganisms isolated from such environments are considered to constitute a valuable tool in the treatment of highly polluted mine water. Knowing that microorganisms, with bacteria in the lead, represent the largest reservoir of undescribed biodiversity (Tekere et al. 2011); their study has proved to be of great importance to science. This study aims at assessing the resistance and bioremediation ability of indigenous bacterial isolate to both V5+ and Ni2+ in a modified wastewater mixed liquor.

Methods

The *Marinobacter* species was isolated according to Huu *et al.* (1999) using Halomonas elongata (HMC) medium. Prior to be used, the isolate was identified by sequencing thereafter exposed to V^{5+} and Ni²⁺ ions at various concentration (from 50 to 800 mg/L) in the modified mixed liquor (Kamika and Momba 2011). Prior to assessing the metal tolerance ability of the isolates, the optimum growth temperature of MWI-1 was determined by incubating the isolates at various temperatures (25 °C, 30 °C and 35 °C) and in the HMC broth. Sample flasks as well as positive controls were inoculated with the isolates (approximately 100 cfu/mL) incubated at 30 °C \pm 2 °C for 4 days. The median lethal concentration (LC_{50}) of the test metal for each of the test microbial isolates was determined as described by previous investigators (Kamika and Momba 2011). The minimum inhibitory concentration (MIC) of the test metal (referring to the smallest concentration of an antimicrobial agent necessary to inhibit growth of microorganisms) was determined according to Shirdam et al. (2006). MIC values were noted when the isolates failed to grow on the plates. After incubation, the microbial isolates were classified as being sensitive or tolerant to Ni²⁺ according to the inhibition of growth cells.

The genera *Marinobacter* were isolated (MWI-1) from the mine water samples and assessed for their possible ability to resist V⁵⁺ and Ni²⁺. The phylogenetic analysis using the neighbour-joining method with a bootstrap

value of 100 replicates indicated that an MWI-1 [AB793286] isolate belonged to the genus *Marinobacter* and was most closely related to *Marinobacter goseongensis* strain En6 [EF660754.1 and NR044340.1] at a similarity of approximately 97 % (fig. 1).

During the study the growth curves of the MWI-1 in a metal-free medium (HMC broth) revealed a lag-phase between time 0 to 2 followed by an exponential phase from time 2 h to 8 h when inoculated at 25 °C, 30 °C and 35 °C with bacterial counts of 7 log CFU/mL, 8 log CFU/mL and 7 log CFU/mL, respectively. At 30 °C, MWI-1 indicated a second exponential growth (10 logCFU/mL) from time 16 h to the end of the experiment, whereas a death phase was observed at 25 °C and 35 °C, repectively.

As MWI-1 grew very well at 30 °C, its tolerance to V^{5+} and Ni^{2+} in the modified mixed liquor was tested at the said temperature and at pH 7.2 ± 0.2. The growth performance of this isolate in the modified mixed liquor containing either V^{5+} or Ni^{2+} or both at two different concentrations in order to highlight the difference of the toxic effect between the two metals (fig. 2).



0.001

Fig. 1 Phylogenetic tree using the neighbour-joining method, constructed and based on the bacterial 16S rRNA gene sequence detected in the present study along with similar sequences detected from the NCBI and RDP databases.



Fig. 2 Growth performance of MWI-1 in a medium containing either V^{5+} (A) or Ni²⁺ (B) or both (B) at 100 mg/L and 200 mg/L, 30 °C, pH 7.2±0.2.

In general, the growth of the isolates decreased with the increases of the metal concentrations. The MWI-1 was able to significantly grow in the presence of V^{5+} at 100 mg/L (9 log CFU/mL) and 200 mg/L (8 log CFU/mL),

while in the presence of Ni²⁺ MWI-1 could only grow at 100 mg/L. Concomitantly, Ni²⁺ toxicity was able to inhibit the growth in all the volume ratios. Statistical evidence revealed a significant difference (p<0.05) in terms of growth performance between the positive controls and those samples treated with Ni²⁺ while no significant difference (p>0.05) was indicated for MWI-1 treated with V⁵⁺. Another significant difference was also noted for positive controls when compared with those samples treated with both metals concomitantly.

A general observation indicated that the MWI-1 isolate was more tolerant to V5+ than to Ni²⁺ in the modified mixed liquor (Table 1). The MWI-1 isolate could resist V⁵⁺ up to 650-700 mg/L (MIC) and only reached 200–250 mg/L in the presence of Ni²⁺. The tolerance or sensitivity of the MWI-1 isolate was also revealed by its ability to remove V⁵⁺ and Ni²⁺ (Table 1 and 2). In the presence of V^{5+} or Ni^{2+} separately, the MWI-1 was able to remove up to 99.95 % of 100 mg/L V⁵⁺ and 86.42 % of 100 mg/L Ni²⁺. When in concomitance with each other (Table 2), the Ni²⁺ toxicity disturbed the removal of V⁵⁺ in the modified mixed liquor. In consortium, none of the metals (V^{5+}/Ni^{2+}) was removed at a percentage of over 30 %, with the exception of V^{5+} (30.15 %) that was removed at a ratio of 1:1 (100 mg/L/100 mg/L).

Due to their abundance, the *Marinobacter* genus of the *Proteobacteria phylum* was isolated in the present study and assessed for their tolerance to and removal of V⁵⁺ and Ni²⁺ in the modified mixed liquor. The isolate (MWI-1) was firstly sequenced and found to be closely related to *Marinobacter goseongensis*.

	Tolerance limit (mg/L)				Percentage removal (%)			
Experi- ment	24h-LC50 (V ⁵⁺)	MIC (V ⁵⁺)	24h-LC50 (Ni ²⁺)	MIC (Ni ²⁺)	100 mg/L V ⁵⁺	200 mg/L V ⁵⁺	100 mg/L Ni ²⁺	200 mg/L Ni ²⁺
1 st	550	700	250	350	99.95	96.38	81.36	47.28
2^{nd}	500	650	200	300	99.98	91.28	86.25	43.95
3^{rd}	500	700	200	350	98.97	94.65	79.64	39.78
	500-550	650-700	200-250	300-350	99.63	94.10	82.42	43.67

Table 1 Tolerance limit and removal ability of MWI-1 in the modified mixed liquor containing V5+ andNi2+ separately.

	100/100 (mg/	L) [V ⁵⁺ /Ni ²⁺ ,	200/100 (mg/l	L) [V ⁵⁺ /Ni ²⁺ ,	100/200 (mg/L) [V ⁵⁺ /Ni ²⁺ ,	
		v/v]		v/v]		v/v]
	V ⁵⁺ (%)	Ni ²⁺ (%)	V ⁵⁺ (%)	Ni ²⁺ (%)	V ⁵⁺ (%)	Ni ²⁺ (%)
1 st	27.05	26.15	16.31	11.07	9.87	3.68
2^{nd}	25.13	21.08	18.05	13.12	7.38	5.28
3 rd	38.27	24.65	17.15	10.24	11.28	9.17
	30.15	23.96	17.17	11.48	9.51	6.04

Table 2 Removal ability of MWI-1 in the modified mixed liquor containing V^{5+} and Ni²⁺ in consortium.

This MWI-1 showed very good growth at 30 °C in the HMC broth, while at 25 °C and 35 °C a prompt die-off was noted when inoculated in a metal-free media. Findings of this study corroborated those of Roh et al. (2008) who reported that the optimum temperature for *Marinobacter goseongensis* sp. nov. should be between 25 °C and 30 °C. When inoculated in modified mixed liquor, culture media containing V⁵⁺ and Ni²⁺, separately or combined and incubated at 30 °C, reveals a significant growth (p<0.05) of the MWI-1 in the media with V^{5+} when compared to the media with Ni²⁺. The MWI-1 bacterial isolate was more tolerant towards V⁵⁺ than towards Ni²⁺ (Table 1). The toxicity of the test metals in MWI-1 appeared to have a relatively negative effect on the metalremoval ability of the test isolate in the modified mixed liquor with V⁵⁺ indicating the highest level of removal (Table 1). It has been reported that bacterial strains can be characterised as being tolerant towards metal such as Ni²⁺, if it is capable of expressing growth at concentrations higher than 100 mg/L of the metal (Gikas 2008).

Zucconi *et al.* (2003) stated that isolating microorganisms from extreme environments represent an appropriate practice to select metal-resistant strains that could be used for metal removal and bioremediation purposes. Since it has been reported that most of the species of the genus *Marinobacter* are halophilic, heterotrophic neutrophiles and living under extreme environmental conditions such as pH and high salinity, they have been isolated from several habitats such as seawater, petroleum refineries, oil-refineies, and so forth (Guo *et al.* 2007). According to Brito *et al.* (2006), the *Marinobacter* species are the best degraders of hydrocarbons and have been associated with the removal of hydrocarbons in seawater. Owing to their ability to grow under extreme habitats, strains found in *Marinobacter* could have the ability to remove metal in the environment. Researchers have reported that several strains of *Marinobacter spp.* such as *Marinobacter aquaeolei* possess iron transport capabilities and are also capable of oxidising iron (Amin *et al.* 2012).

Conclusion

This study revealed that highly contaminated and toxic mine water effluents are a reservoir of novel microbial species which can adequately be used for the removal of metals in highly polluted effluents. The MWI-1 isolate, closely related to Marinobacter goseongensis, demonstrates high tolerance to both V⁵⁺ and Ni²⁺. Further studies on geochemistry and microbial diversity need to be conducted in order to unveil how the chemistry of the effluent from the vanadium mine in South Africa can affect the microbial diversity of the environment. Furthermore, studies carried out on microbial diversity in extreme environments such as mine water are needed in order to isolate novel hyper-tolerant microbial species for the removal of metals.

Acknowledgement

The authors are grateful to the National Research Foundation (NRF) for the funding of this project (Grant number: M590) and to the South African mining industries for allowing the researchers to use their mine water samples.

References

- Adler R.A., Claassen M, Godfrey L, Turton AR (2007) Water, mining, and waste: an historical and economic
- perspective on conflict management in South Africa. Economics of Peace and Security J 2(2): 33–44.
- Amin SA, Green DH, Al Waheed D, Gardes A, Carrano CJ (2012) Iron transport in the genus *Marinobacter*. Biometals 25(1): 135–147.
- Brito EMS, Guyoneaud RM, Goñi-Urriza M, Ranchou-Peyruse A, Verbaere A, Crapez MAC, Wasserman JCSA, Duran R (2006) Characterization of hydrocarbonoclastic bacterial communities from mangrove sediments in Guanabara Bay, Brazil. Res. Microbiol 157 (8): 752–762.
- Gikas P (2008) Single and combined effects of nickel (Ni(II)) and cobalt (Co(II)) ions on activated sludge and on other aerobic microorganisms: A review. J Hazard Mater 159(2–3): 187–203.
- Guo B, Gu J, Ye Y-G, Tang Y-Q, Kida K, Wu X-L (2007) *Marinobacter segnicrescens* sp. nov., a moderate halophile isolated from benthic sediment of the South China Sea. Int J Syst Evol Microbiol 57: 1970– 1974.
- Huu NB, Denner EBM, Ha DTC, Wanner G, Stan-Lotter H (1999) Marinobacter aquaeolei sp. nov., a halophilic bacterium isolated from a Vietnamese oil-producing well. Int J Syst Bacteriol 49: 367–375.
- Imarla T, Hector SB, Deane SM, Rawlings DE (2006) Resistance determinants of a highly arsenic-resistant strain of *Leptospirillum ferriphilum* isolated from a commercial biooxidationtank. Appl Environ Microbiol 72: 2247–2253.
- Johnson DB, Hallberg KB(2003) The microbiology of acidic mine waters. Research in Microbiol 154: 466–473.

- Kamika I, Momba MNB (2011) Comparing the tolerance limits of selected bacterial and protozoan species to nickel in wastewater systems. Sc Total Environ 410: 172–181.
- Oelofse SHH (2009) Mine water pollution–acid mine decant, effluent and treatment: a consideration of key emerging issues that may impact the state of the environment. In: Krishna, C.S. (ed) Mining: Environment and health concerns, 1st edn. The Icfai University Press, India, pp 84–91.
- Roh SW, Quan ZX, Nam YD, Chang HW, Kim KH, Rhee SK, Oh HM, Jeon CO, Yoon JH, Bae JW (2008) Marinobacter goseongensis sp. nov., from seawater. Int J Syst Evol Microbiol 58(Pt 12): 2866–2870.
- Shirdam R, Khanafari A, Tabatabaee A (2006) Cadmium, nickel and vanadium accumulation by three strains of marine bacteria. Iran J Biotechnol 4(3): 180–187.
- Tekere M, Lotter A, Olivier J, Jonker N, Venter S (2011)
 Metagenomic analysis of bacterial diversity of Siloam hot water spring, Limpopo, South Africa.
 African Journal of Biotechnology, 10(78): 18005– 18012.
- Wang J, Yang D, Zhang Y, Shen J, Van Der Gast C, Hahn MW, Wu, Q (2011) Do Patterns of bacterial diversity along salinity gradients differ from those observed for macroorganisms. PLOSOne, 6(11): e27597.
- Zhou M, Liu Y, Zeng G, Li X, Xu, Fan T(2007). Kinetic and equilibrium studies of Cr(VI) biosorption by dead Bacillus licheniformis biomass. World J. Microbiol Biotechnol, 23: 43–48.
- Zucconi L, Ripa C, Alianiello F, Benedetti A, Onofri S (2003) Lead resistance, sorption and accumulation in a *Paecilomyces lilacinus* strain. Biology and Fertility of Soils, 37: 17–22.